- ¹ Infectious Diseases, Public Health Wales, Cardiff, UK
- ² Department of Microbiology, Morriston Hospital, Swansea, UK
- ³ Microbiology Department, Public Health Wales, Cardiff
- ⁴ Health Protection Team, Public Health Wales, Swansea
- ⁵ Medical Directorate General Practice & Revalidation, NHS Wales Health Education and Improvement Wales, Nantgarw, Rhondda Cynon Taff, UK
- ⁶ Cardiff and Vale University Health Board, Public Health Wales, Cardiff

Correspondence to: B Healy brendan.healy@wales.nhs.uk Cite this as: *BMJ* 2022;378:e061402 http://dx.doi.org/10.1136/bmj-2020-061402 Published: 01 July 2022

PRACTICE POINTER

Immunity and infectivity in covid-19

Claire Johnston, ^{1,2} Harriet Hughes, ³ Sion Lingard, ⁴ Stephen Hailey, ⁵ Brendan Healy^{6,2}

What you need to know

- The risk of SARS-CoV-2 transmission is greatest just before symptom onset and in the early symptomatic period
- There is no surrogate marker to determine infectiousness: PCR positivity overestimates the duration of infectivity and can lead to negative consequences such as delayed surgery, delayed access to health care, and blocking of healthcare systems; culture is not practical; and negative lateral flow tests do not equate exactly with non-infectiousness
- Decisions related to transmission risk must take into account all relevant factors, including the overall risk of infection in the community, the individual's ability to comply with prevention measures, their home and work environment, and the risk profile of their likely future close contacts

Understanding how to assess and communicate risk of transmission and immunity against SARS-CoV-2 is important for all healthcare workers. The evolving evidence base regarding infectivity, risk of transmission, risk of reinfection (dependent on circulating variants), and immunity (influenced by post-infection and post-vaccination waning immunity) can make this very challenging.

There are several reasons why individuals with covid-19 and those caring for them are interested to understand whether they are still infectious:

- Individual concern about passing on infection to others
- Healthcare workers to make risk assessment before patient discharge or interventions
- Policy makers to provide risk reduction recommendations.

This article reviews core underlying principles and explains how interpretation of laboratory data—including polymerase chain reaction (PCR), antigen based lateral flow device (LFD), and antibody testing—can support discussions.

When is an individual non-infectious?

There are insufficient data to precisely delineate when an individual is no longer infectious, and the risk is a continuum with considerable inter-person variability. Individual risk assessments will probably always be required (box 1) and will need to take into account the general risk of infection in the community, including risks posed by new variants (box 2).

Box 1: What to consider when a patient asks if they are still infectious

- The reason for the question—Explore the patient's concerns and the specific nature of the inquiry
- The consequences of labelling the individual as "infectious" (psychological, staffing levels, delayed discharge, delayed surgery, etc)
- The consequences of not regarding the individual as potentially infectious
- The risk from this individual relative to the wider community risk
- The results of tests such as PCR, antigen, and antibody (surrogate markers only)
- Discuss infectiousness in terms of levels of risk
- Advise on measures to mitigate that risk (such as cough hygiene, social distancing, mask/face covering (different grades of mask offer different levels of protection), eye protection, hand hygiene)
- Advise that, although patients may have lingering symptoms after infection that are troublesome, these are not indicative of ongoing infection or ongoing infectiousness

Box 2: Example of an individual risk assessment of infectiousness

An immunocompetent individual who had mild disease and has now recovered after seven days asks you when they will no longer be infectious. They work in retail, sing in a choir, and are the main carer of an elderly relative, for whom they do not have a reliable alternative carer. They are worried about passing on infection to their work colleagues, friends in the choir, and their elderly relative. **Advice for the patient**

- We do not have an exact cut-off point for when someone is no longer infectious. However, in one study of people with mild disease no transmissions occurred five days after the onset of symptoms. Analysis of other data has led scientists to conclude that transmission after 10 days is extremely unlikely.
- You can definitely return to your job in retail after 10 days, as per government advice. You are extremely unlikely to be infectious. You are in fact much less of a risk than other people who haven't had the virus yet as, if they get infected, they may be unaware but be in the most infectious stage, which happens early on.
- Even though you are very unlikely to be infectious, you might want to delay returning to the choir, perhaps until after three weeks. This is simply because you can avoid the choir without any significant detriment to anybody, singing is known to increase the risk of transmission, and even though transmission after day 10 is extremely unlikely, the longer the interval since the time of infection the lower the risk. The virus has been cultured in one immunocompetent individual 18 days after symptom

onset, which is why I have suggested three weeks. Similarly, you may decide to delay visiting elderly or vulnerable family members who you don't need to visit because of the very small potential risk.

However, you are the main carer of one elderly relative, and it is
important that you can visit them because there is a risk of harm if
you are not able to look after them. You can resume caring for them,
as you are extremely unlikely to be infectious at this stage. I would
suggest that you pay careful attention to the various preventive
measures (social distancing, mask wearing, and hand hygiene) as an
additional precaution.

Individuals are most infectious in the early stages of the illness, immediately before and shortly after the onset of symptoms.¹ Interventions that target this highest risk period (such as identification and behavioural modification of individuals with early disease) are likely to have the biggest impact in controlling transmission overall. Infectivity and viral load decline from the onset of symptoms.¹² In one study, no transmissions occurred after day five of symptoms even in household contacts.³ In mild to moderate cases, individuals are considered highly unlikely to be infectious beyond 10 days.45 Over-emphasis on the latter stages of recovery (for example, demonstrating PCR negativity in recovering patients) is unlikely to have a significant impact on transmission and can lead to negative unintended consequences, such as delayed surgery, delayed access to health care, and blocking of healthcare systems. It may still have a place in certain circumstances (for example, among immunocompromised patients).

Guidelines worldwide provide recommendations on when it is safe to return to work, broadly based on the likely infectious period.⁶⁻¹¹ These guidelines continue to evolve and can be referenced for up-to-date information. There is no longer a legal requirement in the UK for someone who has covid-19 to self isolate, although it is still recommended.¹² In Wales healthcare workers are advised to self isolate and to return to work when they have two negative lateral flow test results taken 24 hours apart, starting five days after the date of their initial positive test. Those who continue to test positive are advised to continue testing up to day 10. If they are still positive at that point, they are considered unlikely to still be infectious and they can return to work providing they are medically fit.⁹

Patients in hospital are typically kept in isolation for 10 days from the onset of symptoms (14 days for those who are severely immunocompromised); they are then able to stop isolating providing that they have been afebrile for 48 hours and all their symptoms (except for cough and anosmia) have resolved. This can be reduced if they meet these clinical criteria and have two negative lateral flow test results taken 24 hours apart, starting six days after the date of their initial test.¹⁰

International travel and schools are other areas where transmission risk has been scrutinised. In the case of international travel the concern is primarily related to spread of infectious variants with varying degrees of ability to infect vaccinated individuals. There is still potential for global spread of a more virulent variant of SARS-CoV-2. However, the omicron wave has largely tempered those fears for now. In addition, attempts to prevent infiltration of variants through travel restrictions have to date been largely unsuccessful apart from in countries where very strict travel restrictions are put in place before any threat of introduction of the new variant (for example, New Zealand). Risks of transmission in schools need to be balanced against the negative impact on children's mental wellbeing and education, particularly given that most children are at low risk of complications from covid-19.

Are all individuals equally infectious?

Individuals are not equally infectious. Onward transmission varies according to specific host and contact factors and the nature of the exposure (box 3). Transmission is primarily related to direct contact with an infected individual. In one study, transmission rates on trains were highest in those in adjacent seats (attack rate 3.5% (range o to 10.3%)) and increased with time (0.15% per hour) and proximity.¹³ Transmission in passengers who immediately occupied a positive individual's vacated seat occurred in only one out of 1342 cases (0.075%).¹³ Household contacts (11.8%) are more likely than non-household contacts (1.2%) to develop disease.¹⁴

Box 3: Factors associated with increased risk of transmission

Environment

- Indoors
- Poor ventilation
- Crowding
- Close proximity (roughly <2 metres, but transmission is a continuum, the further away the better)
- Shared facilities
- Cold ambient temperature
- Low humidity

Host factors

- Recently infected (highest risk around the time of symptom onset)
- High viral loads
- Severe disease (risk ratios 3.76 (95% Cl 1.1 to 12.76) and 3.99 (1.00 to 15.84) for severe pneumonia and acute respiratory distress syndrome/sepsis³)
- Age
- Comorbidity
- Immunocompromised

Behavioural

- Singing, shouting, chanting
- Coughing and poor cough etiquette
- Sneezing
- Hugging, kissing
- Mask etiquette
- Hand hygiene
- Aerosol generating procedure
- Duration of contact

Viral factors

 Changes in the viral genome have been linked to increased transmissibility (for example, D614G and variant of concern VOC-202012/01, both of which have changes in the spike protein)

Investigations of outbreaks have demonstrated very high attack rates in specific settings.¹⁵⁻¹⁷ These large scale, "super-spreader" events, ¹⁵⁻¹⁷ are characterised by explosive early growth and sustained transmission in later stages, ¹⁸ with 20% of infected individuals triggering 80% of all infections.¹⁹ As transmission is unpredictable and random in nature (stochastic), exercise caution to not over-interpret data from small groups.²⁰

What other factors affect the risk of transmission?

Transmission is influenced by external factors, which should be considered as part of any assessment (box $_3$):

- Prevention measures—masks,²¹ social distancing, vaccination status, hand hygiene, etc
- The activity being undertaken (such as choir)
- The environment (higher risk in crowded or shared facilities and if ventilation is poor)
- The susceptibility and risk of severe disease among contacts.

Individuals are most infectious just before and just after symptom onset. Infectivity decreases thereafter, with transmission after day 10 considered extremely unlikely following mild or moderate disease. Immunocompromised people and those with severe disease are likely to be infectious for a longer, undefined period. Resolution of symptoms is reassuring, signifying development of immunity with likely reduced risk of transmission. Other preventive measures (hand hygiene, mask wearing, social distancing) reduce residual risk further.

What surrogate markers are used to decide on infectivity?

There is currently no ideal surrogate marker for infectiousness (table 1). Viral culture is not a routinely available test in most settings. PCR overestimates the duration of infectiousness but can underestimate risk by virtue of false negative results. Lateral flow devices (LFDs) identify the most infectious individuals reliably but don't detect all infectious individuals. LFDs do not have the same issues of residual positivity as PCR.

Table 1 Surrogate markers of infectiousness											
	Pros	Cons	Overestimate infectivity	Underestimate infectivity	Limits of detection after symptom onset						
Culture	 Confirms presence of intact, viable and potentially infectious virus Surrogate for infectivity 	 Requires category 3 or 4 laboratory Not routinely available Difficult to perform 	Virus deposited into a favourable environment	 Use of a cell line rather than a natural host Delay in inoculation (death of virus in transit) 	 Maximum time 119 days (in an immunocompromised individual)^{22 23} For mild disease 8 days²⁴²⁵ For severe disease 111 days²⁶ 						
PCR	 Virus can be inactivated before processing Requires category 2 laboratory Widely available Fast turnaround time Sensitive Provides a semi-quantitative result 	Will detect viral fragments and/or dead RNA	 Detects viral fragments and/or dead RNA False positive rate unknown – estimate 0.8-4.0%²⁷ 	• False negative rate ~10-30%	 Maximum time 156 days²² 28_30 Median time to a negative upper respiratory tract test 14.5-24 days³⁰ 31 						
Lateral flow test	 Point of care test: results within 20 minutes Can be taken regularly Minimally invasive options available No transport, laboratory infrastructure, validation, or communication of results required Less sensitive than PCR and less likely to overestimate infectivity 	 Inter-user variability False positive results (persistent in some individuals) Lower sensitivity than PCR, may underestimate infectivity 	Detects viral antigen False positive rate varies according to test being used – range 0-7.6% ³²	Lower sensitivity than PCR False negative rate relative to PCR ~65-89%	• Not known						

Culture

Most recommendations are based on viral PCR and culture (table 1). Viral culture confirms the presence of intact, viable, and potentially infectious virus. Although the circumstances required for viral culture are not the same as for transmission, it is considered a reasonable surrogate. In immunocompetent individuals, positive culture beyond day 10 in patients with mild disease is uncommon.⁴ It is more common in those with severe disease.²⁶ 33 -35 Virus has been detected up to day 18 in mild disease, ²⁴ 25 day 111 in severe disease, ²⁶ and day 119 in an immunocompromised individual.²² 23 Individuals may not be very infectious even when culturable virus is present. One individual with severe infection who was culture-positive at day 111 did not cause any secondary infections

despite quarantine termination at three months.²⁶ Also, no infections occurred in 852 contacts exposed to individuals with mild disease after day five.³

Polymerase chain reaction (PCR)

PCR detects the presence of SARS-CoV-2 viral RNA. Previously, guidelines advocated use of PCR as a surrogate for non-infectiousness but studies on viral dynamics have shown that there are several reasons why this is not appropriate.

- PCR can detect non-viable virus and overestimates the duration of infectivity,^{3 28 29 31} with one surveillance study reporting no secondary cases among 790 contacts of 285 "persistently positive" people.³⁶ Relying on PCR as a measure of non-infectiousness may prolong hospital admission and isolation unnecessarily.³⁰
- Results can fluctuate from positive to negative at all stages of infection, can become positive again even after two consecutive negative tests, ²³ 37 ³⁸ can be detected for longer in those with severe infection, ²² ³⁹ and may fluctuate at the level of detection for several weeks.⁴⁰
- Results vary according to sample site (lower respiratory tract samples remaining positive for longer).³⁸

• False negative results can provide false reassurance.³¹

Results can be semi-quantified by the number of cycles required to reach the predetermined positive threshold—the cycle threshold (CT). Low CT values indicate high viral loads (strong positive <25); high CT values (>35) may indicate low viral loads (weak positive). Weak positive results are most common in the very early and late stages of infection but may also be false positives.⁴¹ The CT value is probably linked to infectiousness^{30 42}; supported by decreased ability to culture the virus as the CT value increases^{4 5 25 33 42} and as found with other diseases.⁴³ The CT value is affected by some external factors, such as swab quality and disease stage (lower in early disease but may be rising), so results need to be interpreted with caution.

Lateral flow devices (LFDs)

LFD antigen tests detect a protein antigen which forms part of the viral wall. When present, it is indicative of ongoing replication and therefore the presence of infectious virus. Comparative studies have shown that it is less sensitive than PCR, detecting around 65-89% of PCR-positive samples.⁴⁴ However, the sensitivity is higher in those with higher viral loads (96% for >1 000 000 copies per mL, 92% for 10 000–1 000 000 copies per mL, and 43% for <10 000 copies per mL⁴⁵) and those who were culture positive (>95%).⁴⁴ It has been estimated that LFD tests would detect 83-89% of cases with PCR-positive contacts.⁴⁶ The rapid turnaround time and practicality of lateral flow tests mean they provide a reasonable testing strategy for reducing infection risk in certain circumstances—such as when PCR testing is not practical, when the consequences of a false negative result are acceptable, and when the balance of risks (immediate LFD result *v* delayed PCR result) favours their use.

When are individuals considered to be immune?

Individuals are understandably keen to know whether they are susceptible to reinfection. Reinfection with phylogenetically distinct variants of SARS-CoV-2 has been reported after as little as 48 days⁴⁷ in an otherwise healthy 25 year old man. Asymptomatic reinfection (PCR positivity)^{48 49} and infection with milder disease^{50 51} and more severe disease^{47 51} have all been described. Over time, infection and reinfection have resulted in milder disease at the population level, which is probably related to improved immunity combined with reduced virulence of emerging strains. Reinfection is more likely to be established in individuals with symptoms and more severe disease. The risk of reinfection is a function of the level of immunity present and the infecting viral strain (for example, vaccine escape

variants), which is in turn dependent on the strain(s) circulating in the community at that time. Immunity decreases with time from infection or vaccination. Reinfection is more likely when a new strain emerges, particularly if that strain has properties that enable it to evade immunity developed from previous infection or vaccination. An example of this was seen with the rapid spread of the omicron variant in late 2021.

What factors can you discuss when asked by a patient if they are immune?

- What is known about the response to SARS-CoV-2 (i.e. immunity lasts at least 90 days and likely longer in most people)
- The different types of immunity (T cell and antibody)
- That current tests are only surrogate markers for immunity and do not take account of immune memory
- Reinfections can occur
- Reinfections are often milder than the first episode
- Recovered individuals should comply with prevention measures to avoid reinfection

Most people will be protected from symptomatic reinfection for at least five months, and the immediate risk of reinfection is low (0.02%, incidence rate 0.36 per 10 000 person weeks).^{52 53} There is evidence of increased protection from infection in individuals who are vaccinated after a primary infection, with one prospective cohort study showing that infection-acquired immunity waned after one year in unvaccinated participants but remained consistently higher than 90% in those who were subsequently vaccinated, even in people infected more than 18 months previously.⁵⁴

Immunity in coronavirus infections

Evidence from infections with other coronaviruses (seasonal coronaviruses, MERS-CoV, SARS-CoV-1) and surrogate markers of immunity (antibody and T cell responses) can help inform our understanding of immunity in SARS-CoV-2

Seasonal coronavirus

Serological studies from the 1960s suggest cycling of infection, with different coronavirus strains predominating every two to four years.⁵⁵ Re-challenge experiments (table 2) suggest complete immunity from symptomatic reinfection for at least one year if "reinfected" with the same strain, but only partial immunity when exposed to a heterologous strain.^{56 57} Short duration asymptomatic shedding is possible following re-challenge with the same strain.⁶⁰

Table 2 | Coronavirus re-challenge experiments

Study	Subject	Strain	Time between exposure	No of participants	Antibody positive before re-challenge	Outcome
Reed 1984 ⁵⁶ *	Human	Identical	8-12 months	6	5/6	6/6 completely immune
	-	Heterologous	12 months	12	_	7/12 developed cold symptoms 8/12 shed virus for a short period, including one asymptomatic
Callow 1990 ⁵⁷ †	Human	Identical	12 months	9	7/9	No clinical symptoms 6/9 shed virus for a short period
Chandrashekar 2020 ⁵⁸	Rhesus monkeys	SARS-CoV-2	35 days	9	9/9 neutralising antibodies. Boosted on re-challenge in all	Limited RNA in BAL of 3/9 Low levels of sub-genomic messenger RNA in 4/9
Corbett 2020 ⁵⁹	Rhesus monkeys vaccinated	SARS-CoV-2	4 weeks after second vaccine	8 low dose 7 high dose	15/15 neutralising antibodies	15/15 reduced lung infection Reduced shedding, particularly in high dose group

* Full immunity to the same strain lasts at least one year but only partial immunity is present when exposed to a heterologous strain.5

[†] Some short duration asymptomatic shedding possible on re-challenge/reinfection.57

Immunity to seasonal coronavirus is not lifelong.⁶⁰ Most children are seropositive for seasonal coronavirus by age 3.5 years, yet seasonal coronavirus infections account for ~25% of acute respiratory illness into adulthood.⁶⁰

SARS-CoV-2

Data on immune response to infection and vaccination is continuing to evolve. Presence of antibody is not proof of immunity. Neutralising antibody tests are considered most predictive of protection but are not available routinely. Neutralising antibodies develop in most infected individuals (>90%),⁶¹ although in some the levels are very low or absent,⁶² suggesting that other elements of the immune system are driving recovery.

Antibody responses are stronger and last longer after severe infection.⁶³ Given the protective nature of antibodies in seasonal coronavirus infection, we might expect protection against the same strain to last for most people for at least 12 months. However, viral evolution may be more frequent and common in the early phases of the pandemic, and immunity akin to that seen in studies of seasonal coronavirus in adults may take time to develop.

There are currently four approved vaccines in the UK⁶⁴ and more available worldwide. Data from vaccination studies show that protection wanes over time but lasts in most people for at least four months.⁶⁵ Pfizer vaccine was effective against symptomatic disease in 96% up to two months, 90% for two to four months, and 84% for four to six months. Protective efficacy of the vaccine from symptomatic disease varies according to viral strain and patient age in the range of 70%⁶⁶ to 95%.⁶⁵ Protection against severe infection, hospitalisation, and death is higher still. At the time of writing vaccination has proved effective against all naturally circulating strains. Evidence regarding the efficacy against the latest variant (omicron) is continuing to emerge, although protection is definitely reduced.⁶⁷ Immunity derived from vaccination declines over time. In recognition of this, the UK Joint Committee on Vaccination and Immunisation has recommended a fourth vaccine dose (Spring booster approximately six months after the previous dose) for those at higher risk of covid-19. This will likely be repeated

in the autumn. Decisions on booster vaccinations for the general population will be made in response to evolving evidence.

In summary, infection with coronaviruses do not result in lifelong immunity, and reinfection is common. The natural course for coronavirus infection includes repeat exposure and repeat infection over a variable time course. Over time, SARS-CoV-2 will likely transform into a seasonal coronavirus infection. With the development of increased immunity the risk of re-exposure and reinfection will decline, and the period between episodes will likely increase.

Patient's perspective: Uncertainty about immunity

When I discovered I had covid-19, I had numerous symptoms and remained unwell for a protracted period. The symptoms lifted suddenly after five and a half months when I woke up feeling better.

When I started to go out, I was extremely cautious despite it being weeks since the onset of my symptoms. I was conscious of not touching any walls—what if an elderly person touched the same wall hours later, caught the virus from me, and died? If someone walked down the street, I gave them a wide berth. I questioned whether it was irresponsible of me to leave the house for a walk on my road—I checked and double-checked the guidance.

I still have mixed feelings about how information on immunity affects my decision making. It is now six months since the onset of my symptoms. Part of my confidence in visiting vulnerable relatives comes from a sense that I am less likely to pass covid on to them unknowingly because I am less likely to get infected again. But then I worry about reinfections—what if I get covid again but have very few symptoms and unknowingly spread it? The uncertainty about immunity makes some decision making hard—who to see and when. What happens when our immunity runs out? And will I ever know when this happens? I do not feel that having had covid removes much of this uncertainty. It hasn't really added much confidence for me, as I still have so many unanswered questions.

Additional educational resources

Centers for Disease Control and Prevention

PRACTICE

- COVID-19: Ending isolation and precautions for people with COVID-19: interim guidance. 2022. https://www.cdc.gov/coronavirus/2019ncov/hcp/disposition-in-home-patients.html
- COVID-19: Interim infection prevention and control recommendations for healthcare personnel during the coronavirus disease 2019 (COVID-19) pandemic. 2022. https://www.cdc.gov/coronavirus/2019ncov/hcp/disposition-hospitalized-patients.html
- Interim guidance for managing healthcare personnel with SARS-CoV-2 infection or exposure to SARS-CoV-2. 2022. https://www.cdc.gov/coronavirus/2019-ncov/hcp/guidance-riskassesment-hcp.html

European Centre for Disease Prevention and Control

- Guidance on ending the isolation period for people with COVID-19third update. 2022. https://www.ecdc.europa.eu/en/publicationsdata/covid-19-guidance-discharge-and-ending-isolation
- Contact tracing in the European Union: public health management of persons, including healthcare workers, who have had contact with covid-19 cases – fourth update. 2021. https://www.ecdc.europa.eu/en/covid-19-contact-tracing-public-health-management

UK Health Security Agency

 COVID-19: the green book, chapter 14a. Coronavirus (COVID-19) vaccination information for public health professionals. 2022. https://www.gov.uk/government/publications/covid-19-the-greenbook-chapter-14a

Information resources for patients

UK Health Security Agency

- Coronavirus (COVID-19): guidance for the public. 2022. https://www.gov.uk/government/collections/coronavirus-covid-19list-of-guidance#guidance-for-the-public
- Coronavirus (COVID-19): antibody testing. 2022. https://www.gov.uk/government/publications/coronavirus-covid-19-antibody-tests/coronavirus-covid-19-antibody-tests
- COVID-19: guidance on protecting people defined on medical grounds as extremely vulnerable. 2022. https://www.gov.uk/government/publications/guidance-on-shielding-and-protecting-extremely-vulnerablepersons-from-covid-19

NHS

- Changes to testing for coronavirus (COVID-19). 2022. https://www.nhs.uk/conditions/coronavirus-covid-19/testing/gettested-for-coronavirus/
- National infection prevention and control manual for England. 2022. https://www.england.nhs.uk/publication/national-infection-prevention-and-control/

Questions for future research

- How long are individuals with covid-19, particularly those infected with new strains, infectious for?
- Is there a reliable surrogate marker for infectiousness?
- Can the cycle threshold (CT) value of PCR tests (or quantitative PCR) be used to predict non-infectiousness?
- How are lateral flow and PCR tests best deployed to aid risk decision making?
- Is there a reliable surrogate marker for immunity that predicts protection from reinfection or significant illness?

Education into practice

 How would you discuss the uncertainty around immunity with your patients?

- How do you use viral detection tests (PCR, lateral flow, and other viral antigen tests) when discussing risk of transmission with patients?
- Reflect on a recent case of covid-19 where the individual was worried about onward transmission or duration of immunity? Would you do anything differently having read this article?

How this article was created

This article was assembled using the expertise of the writing group to appraise the key parts of evidence in each heading. A targeted Medline search was carried out for SARS-CoV-2 culture on 5 December 2021. It yielded 3793 results. Titles were screened for papers that discussed culture of SARS-Co-V-2. Relevant abstracts were reviewed, and full articles reviewed where appropriate. Three review articles with more extensive search criteria were scrutinised for data relevant to this article. The guideline section was constructed after accessing guidelines from major European countries, UKHSA, ECDC, CDC, and WHO on 5 December 2021.

How patients were involved in the creation of this article

The article was reviewed by two patients. Their opinions were used to guide the focus of the article and to respond to main concerns. Also one patient wrote a perspective to highlight the considerations and concerns that a patient may have.

Contributors: BH and CJ produced the article first draft. The article was reviewed, edited, and rewritten by HH (microbiology advice including expertise in compiling guidelines for use in Wales), SL (public health advice including expertise in advising and managing cases in the community and discharge from hospital into the community), and SH (primary care advice).

Competing interests: We have read and understood *BMJ* policy on declaration of interests and have no relevant interests to declare.

Provenance and peer review: Commissioned, based on an idea by the authors; externally peer reviewed.

- ¹ World Health Organization. Transmission of SARS-CoV-2: implications for infection prevention precautions: scientific brief. 2020. https://apps.who.int/iris/handle/10665/333114.
- 2 He X, Lau EHY, Wu P, etal. Temporal dynamics in viral shedding and transmissibility of COVID-19. Nat Med 2020;26:-5. . doi: 10.1038/s41591-020-0869-5 pmid: 32296168
- ³ Cheng HY, Jian SW, Liu DP, Ng TC, Huang WT, Lin HHTaiwan COVID-19 Outbreak Investigation Team. Contact tracing assessment of COVID-19 transmission dynamics in Taiwan and risk at different exposure periods before and after symptom onset. *JAMA Intern Med* 2020;180:-63. doi: 10.1001/jamainternmed.2020.2020 pmid: 32356867
- 4 Walsh KA, Spillane S, Comber L, etal. The duration of infectiousness of individuals infected with SARS-CoV-2. J Infect 2020;81:-56. . doi: 10.1016/j.jinf.2020.10.009 pmid: 33049331
- 5 Cevik M, Tate M, Lloyd O, etal. SARS-CoV-2, SARS-CoV-1, and MERS-CoV viral load dynamics, duration of viral shedding, and infectiousness: a living systematic review and meta-analysis. *Lancet* 2020; doi: 10.2139/ssrn.3677918.
- 6 World Health Organization. Criteria for releasing COVID-19 patients from isolation: scientific brief. 2020. https://www.who.int/publications/i/item/criteria-for-releasing-covid-19-patients-from-isolation
- 7 Centers for Disease Control and Prevention. Ending isolation and precautions for people with COVID-19: interim guidance. 2022. https://www.cdc.gov/coronavirus/2019-ncov/hcp/durationisolation.html.
- 8 European Centre for Disease Prevention and Control. Guidance on ending the isolation period for people with COVID-19, third update. 2022. https://www.ecdc.europa.eu/en/publications-data/covid-19-guidance-discharge-and-ending-isolation.
- 9 Welsh Government. Self-isolation: guidance for people with COVID-19 and their contacts. 2022. https://gov.wales/self-isolation.
- 10 Public Health Wales. Infection prevention and control measures for SARS-CoV-2 (COVID-19) in health and care settings - Wales. 2022. https://phw.nhs.wales/services-and-teams/harp/infectionprevention-and-control/guidance/infection-prevention-and-control-measures-for-sars-cov-2covid-19-in-health-and-care-settings-wales/.
- Park M, Pawliuk C, Nguyen T, etal. Determining the period of communicability of SARS-CoV-2: a rapid review of the literature, 2020. *medRxiv* 2020. 20163873, doi: 10.1101/2020.07.28.20163873.
- NHS UK. What to do if you have coronavirus (COVID-19) or symptoms of COVID-19. 2022. https://www.nhs.uk/conditions/coronavirus-covid-19/self-isolation-and-treatment/when-to-self-isolate-and-what-to-do/.
- ¹³ Hu M, Lin H, Wang J, etal. Risk of coronavirus disease 2019 transmission in train passengers: an epidemiological and modeling study. *Clin Infect Dis* 2021;72:-10. doi: 10.1093/cid/ciaa1057 pmid: 32726405

- ¹⁴ Park YJ, Choe YJ, Park O, etalCOVID-19 National Emergency Response Center, Epidemiology and Case Management Team. Contact tracing during coronavirus disease outbreak, South Korea, 2020. *Emerg Infect Dis* 2020;26:-8. doi: 10.3201/eid2610.201315 pmid: 32673193
- ¹⁵ James A, Eagle L, Phillips C, etal. High COVID-19 attack rate among attendees at events at a church - Arkansas, March 2020. *MMWR Morb Mortal Wkly Rep* 2020;69:-5. . doi: 10.15585/mmwr.mm6920e2 pmid: 32437338
- ¹⁶ Hamner L, Dubbel P, Capron I, etal. High SARS-CoV-2 attack rate following exposure at a choir practice - Skagit County, Washington, March 2020. *MMWR Morb Mortal Wkly Rep* 2020;69:-10. doi: 10.15585/mmwr.mm6919e6 pmid: 32407303
- 17 Szablewski CM, Chang KT, Brown MM, etal. SARS-CoV-2 transmission and infection among attendees of an overnight camp - Georgia, June 2020. *MMWR Morb Mortal Wkly Rep* 2020;69:-5. . doi: 10.15585/mmwr.mm6931e1 pmid: 32759921
- ¹⁸ Frieden TR, Lee CT. Identifying and interrupting superspreading events-implications for control of severe acute respiratory syndrome coronavirus 2. *Emerg Infect Dis* 2020;26:-66. doi: 10.3201/eid2606.200495 pmid: 32187007
- 19 Stein RA. Super-spreaders in infectious diseases. Int J Infect Dis 2011;15:-3. doi: 10.1016/j.ijid.2010.06.020 pmid: 21737332
- 20 Wong SCY, Kwong RT, Wu TC, etal. Risk of nosocomial transmission of coronavirus disease 2019: an experience in a general ward setting in Hong Kong. J Hosp Infect 2020;105:-27. . doi: 10.1016/j.jhin.2020.03.036 pmid: 32259546
- 21 World Health Organization. Advice on the use of masks in the context of COVID-19: interim guidance, 6 April 2020. 2020. https://apps.who.int/iris/handle/10665/331693.
- 22 Baang JH, Smith C, Mirabelli C, etal. Prolonged severe acute respiratory syndrome coronavirus 2 replication in an immunocompromised patient. *J Infect Dis* 2021;223:-7. . doi: 10.1093/infdis/jiaa666 pmid: 33089317
- 23 Aydillo T, Gonzalez-Reiche AS, Aslam S, etal. Shedding of viable SARS-CoV-2 after immunosuppressive therapy for cancer. *N Engl J Med* 2020;383:-8. . doi: 10.1056/NEJMc2031670 pmid: 33259154
- 24 Liu WD, Chang SY, Wang JT, etal. Prolonged virus shedding even after seroconversion in a patient with COVID-19. J Infect 2020;81:-56. doi: 10.1016/j.jinf.2020.03.063 pmid: 32283147
- 25 Basile K, McPhie K, Carter I, etal. Cell-based culture informs infectivity and safe de-isolation assessments in patients with coronavirus disease 2019. *Clin Infect Dis* 2021;73:-9. . doi: 10.1093/cid/ciaa1579 pmid: 33098412
- 26 Abe T, Ikeda T, Tokuda Y, etal. A patient infected with SARS-CoV-2 over 100 days. Q/M 2021;114:-9. . doi: 10.1093/qjmed/hcaa296 pmid: 33064816
- 27 Surkova E, Nikolayevskyy V, Drobniewski F. False-positive COVID-19 results: hidden problems and costs. *Lancet Respir Med* 2020;8:-8. doi: 10.1016/S2213-2600(20)30453-7 pmid: 33007240
- ²⁸ Wu J, Liu X, Liu J, etal. Coronavirus disease 2019 test results after clinical recovery and hospital discharge among patients in China. *JAMA Netw Open* 2020;3:e209759.. doi: 10.1001/jamanetworkopen.2020.9759 pmid: 32442288
- ²⁹ Xiao AT, Tong YX, Gao C, Zhu L, Zhang YJ, Zhang S. Dynamic profile of RT-PCR findings from 301 COVID-19 patients in Wuhan, China: a descriptive study. *J Clin Virol* 2020;127:104346. . doi: 10.1016/j.jcv.2020.104346 pmid: 32361324
- 30 Walsh KA, Jordan K, Clyne B, etal. SARS-CoV-2 detection, viral load and infectivity over the course of an infection. J Infect 2020;81:-71. doi: 10.1016/j.jinf.2020.06.067 pmid: 32615199
- 31 Gombar S, Chang M, Hogan CA, etal. Persistent detection of SARS-CoV-2 RNA in patients and healthcare workers with COVID-19. *J Clin Virol* 2020;129:104477. . doi: 10.1016/j.jcv.2020.104477 pmid: 32505778
- 32 Mistry DA, Wang JY, Moeser ME, Starkey T, Lee LYW. A systematic review of the sensitivity and specificity of lateral flow devices in the detection of SARS-CoV-2. *BMC Infect Dis* 2021;21: doi: 10.1186/s12879-021-06528-3 pmid: 34407759
- 33 Gniazdowski V, Paul Morris C, Wohl S, etal. Repeated coronavirus disease 2019 molecular testing: correlation of severe acute respiratory syndrome coronavirus 2 culture with molecular assays and cycle thresholds. *Clin Infect Dis* 2021;73:-9. . doi: 10.1093/cid/ciaa1616 pmid: 33104776
- ³⁴ van Kampen JJA, van de Vijver DAMC, Fraaij PLA, etal. Duration and key determinants of infectious virus shedding in hospitalized patients with coronavirus disease-2019 (COVID-19). *Nat Commun* 2021;12:. doi: 10.1038/s41467-020-20568-4 pmid: 33431879
- ³⁵ Folgueira MD, Luczkowiak J, Lasala F, Perez-Rivilla A, Delgado R. Persistent SARS-CoV-2 replication in severe COVID-19.*medRxiv* (2020).
- 36 Korea Centers for Disease Control and Prevention. Findings from Investigation and analysis of re-positive cases. 2020. https://www.mofa.go.kr/eng/brd/m_22743/view.do?seq=3&srch-Fr&%3BsrchTo&%3BsrchWord&%3BsrchTp&%3Bmulti_itm_seq=0&%3Bitm_seq_1=0&%3Bitm_seq_2=0&%3Bcompany_cd&%3Bcompany_nm&page=1&titleNm.
- 37 Chen D, Xu W, Lei Z, etal. Recurrence of positive SARS-CoV-2 RNA in COVID-19: A case report. Int J Infect Dis 2020;93:-9. . doi: 10.1016/j.ijid.2020.03.003 pmid: 32147538
- ³⁸ Lui G, Ling L, Lai CK, etal. Viral dynamics of SARS-CoV-2 across a spectrum of disease severity in COVID-19. *J Infect* 2020;81:-56. doi: 10.1016/j.jinf.2020.04.014 pmid: 32315724
- ³⁹ Zheng S, Fan J, Yu F, etal. Viral load dynamics and disease severity in patients infected with SARS-CoV-2 in Zhejiang province, China, January-March 2020: retrospective cohort study. *BMJ* 2020;369:. doi: 10.1136/bmj.m1443 pmid: 32317267
- 40 Lee S, Kim T, Lee E, etal. Clinical course and molecular viral shedding among asymptomatic and symptomatic patients with SARS-CoV-2 infection in a community treatment center in the Republic of Korea. JAMA Intern Med 2020;180:-52. . doi: 10.1001/jamainternmed.2020.3862 pmid: 32780793

the bmj | BMJ 2022;378:e061402 | doi: 10.1136/bmj-2020-061402

- 41 Healy B, Khan A, Metezai H, Blyth I, Asad H. The impact of false positive COVID-19 results in an area of low prevalence. *Clin Med (Lond)* 2021;21:-6. doi: 10.7861/clinmed.2020-0839 pmid: 33243836
- 42 La Scola B, Le Bideau M, Andreani J, etal. Viral RNA load as determined by cell culture as a management tool for discharge of SARS-CoV-2 patients from infectious disease wards. *Eur J Clin Microbiol Infect Dis* 2020;39:-61. doi: 10.1007/s10096-020-03913-9 pmid: 32342252
- ⁴³ Fløe A, Hilberg O, Thomsen VØ, Lillebaek T, Wejse C. Shortening isolation of patients with suspected tuberculosis by using polymerase chain reaction analysis: A nationwide cross-sectional study. *Clin Infect Dis* 2015;61:-73. doi: 10.1093/cid/civ563 pmid: 26175524
- Pickering S, Batra P, Merrick B, etal. Comparative performance of SARS-CoV-2 lateral flow antigen tests and association with detection of infectious virus in clinical specimens: a single-centre laboratory evaluation study. *Lancet Microbe* 2021;2:-71. doi: 10.1016/S2666-5247(21)00143-9 pmid: 34226893
- 45 Department of Health & Social Care. Key points summary: asymptomatic testing for SARS-CoV-2 using antigen-detecting lateral flow devices (evidence from performance data October 2020 to May 2021). 2021. https://www.gov.uk/government/publications/lateral-flow-device-performancedata/key-points-summary-asymptomatic-testing-for-sars-cov-2-using-antigen-detecting-lateralflow-devices-evidence-from-performance-data-october-2020-to-m.
- 46 Lee LYW, Rozmanowski S, Pang M, etal. Severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) infectivity by viral load, S gene variants and demographic factors, and the utility of lateral flow devices to prevent transmission. *Clin Infect Dis* 2022;74:-15. . doi: 10.1093/cid/ciab421 pmid: 33972994
- 47 Tillett R, Sevinsky J, Hartley P, etal. Genomic evidence for a case of reinfection with SARS-CoV-2.SSRN. 2020doi: 10.2139/ssrn.3680955.
- 48 To KK, Hung IF, Ip JD, etal. Coronavirus disease 2019 (COVID-19) re-infection by a phylogenetically distinct severe acute respiratory syndrome coronavirus 2 strain confirmed by whole genome sequencing. *Clin Infect Dis* 2021;73:-51. doi: 10.1093/cid/ciaa1275 pmid: 32840608
- 49 Gupta V, Bhoyar RC, Jain A, et al. Asymptomatic reinfection in two healthcare workers from India with genetically distinct SARS-CoV-2. 2020. https://osf.io/4fmrg/.
- 50 Lee JS, Kim SY, Kim TS, etal. Evidence of severe acute respiratory syndrome coronavirus 2 reinfection after recovery from mild coronavirus disease 2019. *Clin Infect Dis* 2021;73:-8. doi: 10.1093/cid/ciaa1421 pmid: 33219681
- 51 Sciscent BY, Eisele CD, Ho L, King SD, Jain R, Golamari RR. COVID-19 reinfection: the role of natural immunity, vaccines, and variants. *J Community Hosp Intern Med Perspect* 2021;11:-9. doi: 10.1080/20009666.2021.1974665 pmid: 34804382
- 52 Hall VJ, Foulkes S, Charlett A, etalSIREN Study Group. SARS-CoV-2 infection rates of antibody-positive compared with antibody-negative health-care workers in England: a large, multicentre, prospective cohort study (SIREN). *Lancet* 2021;397:-69. doi: 10.1016/S0140-6736(21)00675-9 pmid: 33844963
- ⁵³ Abu-Raddad LJ, Chemaitelly H, Malek JA, etal. Assessment of the risk of severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) reinfection in an intense reexposure setting. *Clin Infect Dis* 2021;73:-40. . doi: 10.1093/cid/ciaa1846 pmid: 33315061
- ⁵⁴ Hall V, Foulkes S, Insalata F, etalSIREN Study Group. Protection against SARS-CoV-2 after Covid-19 vaccination and previous infection. *N Engl J Med* 2022;386:-20. . doi: 10.1056/NEJMoa2118691 pmid: 35172051
- 55 Monto AS. Medical reviews. Coronaviruses. Yale J Biol Med 1974;47:-51.pmid: 4617423
- 56 Reed SE. The behaviour of recent isolates of human respiratory coronavirus in vitro and in volunteers: evidence of heterogeneity among 229E-related strains. *J Med Virol* 1984;13:-92. . doi: 10.1002/jmv.1890130208 pmid: 6319590
- 57 Callow KA, Parry HF, Sergeant M, Tyrrell DA. The time course of the immune response to experimental coronavirus infection of man. *Epidemiol Infect* 1990;105:-46. . doi: 10.1017/S0950268800048019 pmid: 2170159
- 58 Chandrashekar A, Liu J, Martinot AJ, etal. SARS-CoV-2 infection protects against rechallenge in rhesus macaques. *Science* 2020;369:-7. . doi: 10.1126/science.abc4776 pmid: 32434946
- ⁵⁹ Corbett KS, Flynn B, Foulds KE, etal. Evaluation of the mRNA-1273 Vaccine against SARS-CoV-2 in Nonhuman Primates. *N Engl J Med* 2020;383:-55. . doi: 10.1056/NEJMoa2024671 pmid: 32722908
- 60 Kellam P, Barclay W. The dynamics of humoral immune responses following SARS-CoV-2 infection and the potential for reinfection. *J Gen Virol* 2020;101:-7. . doi: 10.1099/jgv.0.001439 pmid: 32430094
- 61 O Murchu E, Byrne P, Walsh KA, etal. Immune response following infection with SARS-CoV-2 and other coronaviruses: A rapid review. *Rev Med Virol* 2021;31:e2162. doi: 10.1002/rmv.2162 pmid: 32964627
- 62 Kellam OR, Wu F, Wang A, etal. Neutralizing antibody responses to SARS-CoV-2 in a COVID-19 recovered patient cohort and their implications. *medRxiv* 2020.
- ⁶³ Ibarrondo FJ, Fulcher JA, Goodman-Meza D, etal. Rapid decay of anti-SARS-CoV-2 antibodies in persons with mild Covid-19. *N Engl J Med* 2020;383:-7. doi: 10.1056/NEJMc2025179 pmid: 32706954
- 64 UK Health Security Agency. COVID-19: the green book, chapter 14a. Coronavirus (COVID-19) vaccination information for public health professionals. 2022. https://www.gov.uk/government/publications/covid-19-the-green-book-chapter-14a
- ⁶⁵ Polack FP, Thomas SJ, Kitchin N, etalC4591001 Clinical Trial Group. Safety and Efficacy of the BNT162b2 mRNA Covid-19 Vaccine. N Engl J Med 2020;383:-15. . doi: 10.1056/NEJMoa2034577 pmid: 33301246

7

- Voysey M, Clemens SAC, Madhi SA, etalOxford COVID Vaccine Trial Group. Safety and efficacy of the ChAdOx1 nCoV-19 vaccine (AZD1222) against SARS-CoV-2: an interim analysis of four randomised controlled trials in Brazil, South Africa, and the UK. *Lancet* 2021;397:-111. doi: 10.1016/S0140-6736(20)32661-1 pmid: 33306989
- 67 Andrews N, Stowe J, Kirsebom F, etal. Covid-19 vaccine effectiveness against the omicron (B.1.1.529) variant. N Engl J Med 2022;386:-46. doi: 10.1056/NEJMoa2119451 pmid: 35249272