visual and auditory effects. We also thank Drs S F Damluji, L Magos, and D O Marsh for reading and criticising this paper.

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Lichenoid tattoo hypersensitivity

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Summary and conclusions

Four patients are described who developed granulomatous reactions in the red portions of their tattoos. Histopathological and immunofluorescence studies showed features of lichen planus. Mercury was identified in only one patient's lesion, and hypersensitivity to mercury was shown by patch testing in one other patient. Tattooing may provide a localised antigenic challenge resulting in spontaneous occurring lichen planus.

Introduction

Tattoos occasionally have intriguing and even disastrous consequences in addition to remorse. Fortunately most complications of tattoos are rare, but of the complications that do occur a granulomatous response in the red portion is the most common. We describe four patients who presented over a six-month period with tattoo granuloma who had the same unusual tissue reaction.

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Patients

The main clinical features of the four patients at presentation are shown in the table. The affected areas of all the tattoos were raised up to 0.5 cm above the level of the surrounding skin, were sharply confined to the red areas, and were slightly scaly or crusty in places (see fig 1). Three patients (cases 1, 2, and 4) were tattooed by the same tattooist in Cardiff within a nine-month period in 1976-7. The other patient (case 3) could not remember which tattooist was responsible for which tattoo, but knew that they were all done in Cardiff. The tattooist in cases 1, 2, and 4 was questioned about his technique. Samples of the pigments that he said had been used were obtained for analysis.

Investigations and results

Histological findings—Biopsy specimens of the swollen red portions of all four patients' tattoos were examined histologically and by immunofluorescence. In all cases the histological features were similar. Scattered throughout the dermis were irregular granulomas and clumps of an amorphous black material that was assumed to be tattoo pigment. Most of the pigment was extracellular and did not seem to be related to the inflammatory cell infiltrate or to the degree of lichenoid change. The inflammatory cell infiltrate was dense and not aggregated around any anatomical structure. Lymphocytes and histiocytes predominated but there were also polymorphonuclear leucocytes and plasma cells. The inflamed areas were oedematous and there were small collections of extravasated red blood cells. The overlying epidermis showed quite striking lichenoid features (fig 2). There was irregular hyperkeratosis and hyperkeratosis interspersed with areas of epidermal atrophy. The dermoepidermal junction showed the typical “sawtooth” profile of lichen planus in some sites. The normal basal layer of the epidermis was replaced by a patchily vacuolated layer of cells that showed pronounced cytoid-body formation and was infiltrated in places by inflammatory cells. Cytoid bodies were grouped in grape-like clumps below the dermoepidermal junction (fig 3).

Immunofluorescence—Cryostat sections were examined by direct immunofluorescence with a Nikon microscope fitted with a
Clinical features at presentation of four patients with granulomatous reaction to tattooing

<table>
<thead>
<tr>
<th>Case No</th>
<th>Age (years)</th>
<th>Sex</th>
<th>Site of tattoo</th>
<th>Interval between tattoo and reaction</th>
<th>General health</th>
<th>Previous medical history</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>25</td>
<td>M</td>
<td>Right forearm</td>
<td>3 months</td>
<td>Good</td>
<td>Not relevant</td>
</tr>
<tr>
<td>2</td>
<td>23</td>
<td>M</td>
<td>Right forearm</td>
<td>1 month</td>
<td>Good</td>
<td>Not relevant</td>
</tr>
<tr>
<td>3</td>
<td>27</td>
<td>M</td>
<td>Right upper arm</td>
<td>Unknown</td>
<td>Chronic alcoholic</td>
<td>Not relevant</td>
</tr>
<tr>
<td>4</td>
<td>18</td>
<td>M</td>
<td>Right forearm</td>
<td>“A few weeks”</td>
<td>Good</td>
<td>Not relevant</td>
</tr>
</tbody>
</table>

Patient was highly sensitive to nickel sulphate and slightly sensitive to potassium dichromate. The fourth patient showed no sensitivity.

**Haematological and serological tests**—One patient (case 3) had hypochromic anaemia (haemoglobin 11·2–13·4 g/dl) and a persistently raised erythrocyte sedimentation rate (up to 75 mm in the first hour). Results of all haematological and serological tests were normal in the other three patients.

**Electron-probe microanalysis of tissue sections and pigment**—Paraffin sections of the lesions were deparaffinised in xylene, and cryostat sections were stuck to scanning electron-microscope (SEM) stubs, coated with carbon in an Edwards vacuum-coating unit, and examined in a Cambridge stereoscan SEM fitted with an Edax analysis system. Mercury was specifically sought but was detected in small amounts in granular material in the dermis in the section from only one patient (case 3). Samples of all the pigments supplied were crushed.

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**FIG 1**—Case 4. Appearance of inflammation in red zone (R) of tattoo.

**FIG 2**—Case 4. Skin biopsy specimen showing pronounced lichenoid change, with severe degeneration of basal layer and thickening. (H and E. × 80.)

**FIG 3**—Case 4. Photomicrograph of biopsy specimen showing cytoid bodies (arrowed) grouped below dermoepidermal junction. (H and E. × 80.)

**FIG 4**—Case 2. Skin biopsy specimen showing globular immunofluorescence due to IgG. (H and E. × 80.)
into fine powders and stuck to SEM stubs, coated with carbon, and examined in the SEM. The main constituents of the pigments were as follows. Red: sulphur, silicon, and aluminium, and chlorine; green: titanium, silicon, and copper; white: titanium, aluminium, zinc, calcium, and silicon; orange: titanium and chlorine.

Discussion

The development of granulomatous hypersensitivity to a constituent of the red pigment used in tattoos is well documented. Apparently mercapturic acid is often the agent responsible. Nevertheless, mercury now seems to be an infrequently used tattoo pigment.

We can find no previous report of the dramatic lichenoid response seen in our patients. The resemblance to lichen planus was emphasised by the striking immunofluorescence findings, which included a fibrin band and strongly fluorescent cytid bodies with most of the reagents tested. Clearly, three of the patients (cases 1, 2, and 4) did not have lichen planus (or lupos erythematosus) as there were no other lesions on the skin or mucosae. The solitary tattoo reaction in case 3 was identical with that in the other three patients clinically, histologically, and on direct immunofluorescence. Nevertheless, this patient had had discoid lupus erythematosus with lichenoid features. His reaction was not merely a local expression of his lupos erythematosus induced by the trauma of tattooing as it was confined to the red portion of one tattoo. The relation between this patient’s condition and his odd lichenoid reaction must remain doubtful.

These four patients presented within six months of each other, and the consistent presence of the unusual lichenoid response in all the patients’ lesions surely could not have been coincidental. Scutt stated that no more than 10 patients with tattoo granuloma are seen yearly in the United Kingdom. The question therefore arises whether the tattooist had inadvertently included some other substance in the injected material.

Electron-probe microanalysis showed mercury in the dermis in only one section, but the amount detected was small and could have been a “contaminant.” Mercury hypersensitivity on patch testing was found in only one patient (case 1). The red pigment we obtained contained no mercury. The negative patch test results in the other three patients did not necessarily mean that they were not hypersensitive to injected mercury, as their hypersensitivity had been provoked by intracutaneous injection and not epicutaneous challenge.

The lesions of lichen planus may be associated with immune reactions. Possibly in our patients the local antigenic challenge supplied by the unknown constituent of the tattoo pigment induced a local immunological response resulting in true lichen planus. Or perhaps these patients had been inoculated with an infectious agent which caused the local tissue reaction. If this was the case it has intriguing implications for the possible aetiology of spontaneously occurring lichen planus.

We thank Dr Nick Moore, department of crystallography, Birkbeck College, London, for help with the electron-probe microanalyser.

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SIDE EFFECTS OF DRUGS

Incidence of thrombophlebitis with naftidrofuryl

Woodhouse and Edie recently reported an unusually high incidence of thrombophlebitic complications with intravenous infusions of naftidrofuryl (Praxilene), a drug widely used for treating vascular insufficiency. They described 13 naftidrofuryl infusions (200 mg naftidrofuryl in 200 ml dextrose or common salt solution twice daily) given to seven elderly patients, severe rapidly progressing thrombophlebitis being observed on 10 occasions. Other observers disagree with this experience and reported a low incidence of thrombophlebitis when the drug was given in a dose of 200 mg diluted in 500 ml of dextrose.

Survey and results

To determine the frequency of thrombophlebitic complications from the use of naftidrofuryl infusions in Germany we approached 13 angiological centres for information on their experiences with the drug. The results (see table) showed that the incidence of thrombophlebitis in patients given

Details of naftidrofuryl infusions used in 13 centres* in Germany and incidence of thrombophlebitis

<table>
<thead>
<tr>
<th>Centre</th>
<th>No of infusions</th>
<th>Period of use (years)</th>
<th>Average dose per infusion (mg)</th>
<th>Average duration of infusion (hours)</th>
<th>Basic solution used</th>
<th>Incidence of thrombophlebitis</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>15 000</td>
<td>8</td>
<td>800–1600</td>
<td>2–3</td>
<td>500 ml physiological NaCl</td>
<td>Extremely seldom. No serious or ascending thromboses</td>
</tr>
<tr>
<td>2</td>
<td>7 250</td>
<td>5</td>
<td>400–1200</td>
<td>2–4</td>
<td>500 ml physiological NaCl or 500 ml 5% fructose</td>
<td>None</td>
</tr>
<tr>
<td>3</td>
<td>15 500</td>
<td>6</td>
<td>600–1000</td>
<td>1–3</td>
<td>Rhuscomacroides or physiological NaCl</td>
<td>So far about 15 episodes; connection with infusions not certain</td>
</tr>
<tr>
<td>4</td>
<td>4 650</td>
<td>6</td>
<td>200–400</td>
<td>2–4</td>
<td>500 ml 5%, glucose</td>
<td>None</td>
</tr>
<tr>
<td>5</td>
<td>11 760</td>
<td>6</td>
<td>400</td>
<td>1–2</td>
<td>500 ml physiological NaCl</td>
<td>About 2–8%</td>
</tr>
<tr>
<td>6</td>
<td>20 250</td>
<td>8</td>
<td>120–400</td>
<td>—</td>
<td>500 ml NaCl</td>
<td>No obvious association between thrombosis and infusions</td>
</tr>
<tr>
<td>7</td>
<td>8 500</td>
<td>7</td>
<td>200–800</td>
<td>2–4</td>
<td>500 ml 5% fructose</td>
<td>About 0.5% – 1%; less than the usual risk of thrombophlebitis</td>
</tr>
<tr>
<td>8</td>
<td>18 000</td>
<td>5</td>
<td>400–1400</td>
<td>2–4</td>
<td>Physiological NaCl</td>
<td>None</td>
</tr>
<tr>
<td>9</td>
<td>8 000</td>
<td>8</td>
<td>&lt;2000</td>
<td>1–2</td>
<td>Physiological NaCl or NaCl</td>
<td>Local reactions only</td>
</tr>
<tr>
<td>10</td>
<td>11 750</td>
<td>6</td>
<td>400</td>
<td>1–2</td>
<td>500 ml physiological NaCl</td>
<td>About 0.5%</td>
</tr>
<tr>
<td>11</td>
<td>3 500</td>
<td>4</td>
<td>&lt;1200</td>
<td>4</td>
<td>500 ml physiological NaCl</td>
<td>Normal complication rate (3%).</td>
</tr>
<tr>
<td>12</td>
<td>15 000</td>
<td>5</td>
<td>400</td>
<td>11–2</td>
<td>—</td>
<td>Only three clear cases of thrombosis associated with infusion</td>
</tr>
<tr>
<td>13</td>
<td>20 000</td>
<td>7</td>
<td>&lt;1600</td>
<td>—</td>
<td>—</td>
<td></td>
</tr>
</tbody>
</table>

*Details of the centres listed may be obtained from the author.